



Bibliography

Alvarez, O.M., et al. (1983). The healing of superficial skin wounds is stimulated by external electrical current. *J of Investigative Dermatology*, 81:144-48.

Assimacopoulos, D. (1968). Low intensity negative electric current in treatment of ulcers of leg due to chronic venous insufficiency: preliminary report of three cases. *American J of Surgery*, 115:683-687.

Anderson, JC. & Eriksson, C. Piezoelectric properties of dry and wet bone. *Nature* 227:491--492; 1970.

Barron, J.J., W.E. Jacobson, et al: 1985 Treatment of Decubitus Ulcers, : A New Approach. *Minn med* 68:103-106

Bassett CA & Becker RO. Generation of electric potentials by bone in response to mechanical stress. *Science* 1962;137:1063-4

Bassett, CA; Pawluk, R.; Becker, RO. Effect of electric currents on bone in vivo. *Nature* 204:652-654; 1964.

Bertolucci and Grey: Clinical Comparative Study of Microcurrent Electrical Stimulation to Mid-Laser and Placebo Treatment in Degenerative Joint Disease of the Temporomandibular Joint, *Journal of Craniomandibular Practice*, 1995

Summary: 48 patients were divided into three groups, some receiving placebo, some microcurrent and some laser to treat pain of TMJ syndrome. Both microcurrent and laser were found to be significantly more effective than placebo, with laser slightly more effective than microcurrent. The author acknowledges that lasers are not legally sold in the United States for this purpose, and that microcurrent's easy accessibility makes it more practical for practitioners here.

Becker, R.O. (1982). Electrical control systems and regenerative growth. *J of Bioelectricity*, 1(2):239-264

Becker, R.O., G. Shelden. 1985 *The Body Electric*. William Morrow & Co.: New York

Byl, McKenzie et.al.: Pulsed Microampereage Stimulation: A Controlled Study of Healing of Surgically Induced Wounds in Yucatan Pigs, *Physical Therapy*, Volume 74, Number 3/March 1994

Summary: This study failed to confirm the superiority of microcurrent-stimulated wounds in test pigs over controls. A good review of previous studies is given as well as a discussion of which research variables could account for the inconclusive results, which vary from other studies that found microcurrent to be efficacious for wound healing.

Bourguignon, G.J., L.Y.W. Bourguignon. 1987. Electrical Stimulation of Protein and DNA Synthesis in Human Fibroblasts. *FASEB J* 1(15): 398-402

Brighton CT, Black J & Pollack SR. Electrical properties of Bone and Cartilage: Experimental Effects and Clinical Applications. Grune & Stratton Inc. New York NY 1979.

Carey, L.C., & Lepley, D. (1962). Effect of continuous direct electric current on healing wounds. *Surgical Forum*, 13:33-35.

Carley, P.J., S.F. Wainapel: Electrotherapy for Acceleration of Wound Healing: Low Intensity Direct Current *Archives of Physical Medicine and Rehabilitation*, Vol. 66, 443-445 July 1985

Summary: 30 hospital patients with non healing ulcers were divided into two groups, one treated with conventional wound dressings and one with microcurrent stimulation at 300-700 uA. The latter group was given two hour stimulation periods per day. After six weeks of such treatments, the group treated with microcurrents showed a 150-250% faster healing rate, with stronger scar formation, less pain and lessened infection of the treated area

Chakkalakal DA, Lipello L, Shindell RL, Connolly JF. Electrophysiology of direct current stimulation of fracture healing in canine radius. *IEEE Trans Biomed Eng* 1990;37,11:1048-58.

Chen, C.P. Hess. 1987. Calcium Channels in Mouse 3T3 and Human Fiboblasts. *Biophys j* 51:226a

Cheng, et Al: The Effects of Electric Current on ATP Generation, Protein Synthesis, and Membrane Transport in Rat Skin *Clinical Orthopaedics and Related Research*, #171, Nov/Dec. 1982

Summary: These researchers used in vitro slices of rat skin to determine some of the biochemical explanations for accelerated wound healing demonstrated in the above studies. By applying various levels of current to the samples, and then chemically analysing them, they determined that skin treated at currents below 1000 uA showed up to 75% higher amino acids and up to 400% more available ATP than controls, and that skin treated at levels above 1000 uA showed depressed levels of these substances. Often less than non-treated controls

Childers KR. The Effects of Direct Electric Current on Condylar and Mandibular Growth in the Rabbit. Masters Thesis, University of Tennessee, 1990.

Dayton, P.D & Palladino, S.J. (1989). Electrical stimulation of cutaneous ulcerations: a literature review. *J of the American Podiatric Medical Association*, 79:318-321.

Davidovitch Z, Finkelson M, Steigman S, Shanfield J, Montgomery P & Korostoff E. Electric currents, bone remodelling, and orthodontic tooth movement. The effect of electric currents on periodontal cyclic nucleotides. *AM J Orthod* 1980a;77:14-32.

Davitovitch Z, Finkelson M, Steigman S, Shanfield J, Montgomery P & Korostaff E. Electric currents, bone remodelling, and orthodontic tooth movement. II Increase in the rate of tooth movement and periodontal cyclic nucleotide by combined force and electric currents. *AM J Orthod* 1980b;77:33-47.

Dealler, S. F. Electrical phenomena associated with bones and fractures and the therapeutic use of electricity in fracture healing. *J. Meal. Engin. Technol.* 5:73-79; 1981.

DuPont: Trigger Point Identification and Treatment with Microcurrent, *The Journal of Craniomandibular Practice*, October 1999, Vol. 17, #4

Summary: This article gives the author's techniques for locating and stimulating trigger points (TP's) using a microcurrent stimulator, specifically for the treatment of temporomandibular disorders. He states that electrical conductivity is highest over trigger points, and galvanic skin response (GSR) testing can be used to locate such points. He utilizes probe electrodes to treat small TP's, and pad electrodes to treat larger ones. Probe treatment is delivered @ 0.3 Hz, 20 – 40 uA, with treatment time of 10 – 30 seconds per site. He suggests administering treatment in 24-48 intervals, and states that results should be seen within 2 – 3 treatments. He acknowledges that these protocols are not necessarily the best ones, but work well for his practice.

Eriksson. C Electrical properties of bone. In Bourne GH (ed.): *Biochemistry and Physiology of Bone*. vol. 4. New York. Academic Press, 1976, pp. 329-384.

Falanga, V et al. (1987). Electrical stimulation increases the expression of fibroblast receptors for transforming growth factor-beta. (abstracted). *J of Investigative Dermatology*, 88:488.

Ferrier J, Ross SM, Kanehisa J, Aubin JE. Osteoclasts and osteoblasts migrate in opposite directions in response to a contact electrical field. *J Cell Physiol* 1986;129:283-8.

Firzsimmons RJ, Strong D, Mohan S & Baylink D. Low-amplitude, low-frequency electric field-stimulated bone cell proliferation may in part be mediated by increased IGF-II release. *J Cell Physiol* 1992;150,1:84-9.

Friedenberg ZB, Andrews ET, Smolenski BI, Pearl BW, Brighton CT. Bone reaction to varying amounts of direct current. *Surg. Gynec Obstet* 1970;131:894-9.

Friedenberg ZB, Brighton CT. Bioelectric potentials in bone. *J Bone Joint Surg* 1966;48-A:915-23.

Friedenberg ZB, Harlow MC, Brighton CT. Healing on non-union of the medial malleolus by means of direct current: a case report. *J Trauma* 1971b; 11:883-5

Friedenberg ZB & Kohanim M. Effect of direct current on bone. *Surg Gynec Obstet* 1968;127:97.

Friedenberg, ZB.; Harlow, MC.; Heppenstali, R. B.; Brighton, C. T. The cellular origin of bioelectric potentials in bone. *Calc.Tiss. Res.* 13:53; 1973.

Friedenberg, ZB.; Roberts, P. G.; Didizian, N.H.; Brighton, C. T. Stimulation of fracture healing by direct current in the rabbit fibula. *J. Bone Joint Surg.* 53A: 1400-1408; 1971b.

Friedenberg, ZB.; Zemsky, L. M.; Pollis, R. P.; Brighton, C. T. The response of non-traumatized bone to direct current. *J. Bone Joint Surg.* 56A: 1023-1040; 1974.

Fukada E, Lang S, Mascarenhas S, Pilla A, Shamos M. The electrophysical and electrochemical properties of living tissue. *Ann NY Acad Sci* 1974; 238:228-36.

Fukada E, Yasuda I. On the piezoelectric effect on bone. *J Phys Soc Japan* 1957;12:1158-62.

Gault and Gatens: Use of Low Intensity Direct Current in Management of Ischemic Skin Ulcers Physical Therapy, Vol. 56, #3, March 1976.

Summary: 100 patients with skin ulcers were treated with microcurrent stimulation; six of them had bacterial ulcers with one side used as controls. Stimulation of 200-800 uA was applied, with negative polarity used until infection cleared, and then polarity reversed. Patients had diagnosis ranging from quadriplegia, CVA, brain tumour, peripheral vascular disease, burns, diabetes, fracture, and amputation. The lesions with patients treated with currents showed approximately twice as fast a healing rate.

Goh JC, Bose K, Kang YK, Nugroho B. Effects of electrical stimulation on the biomechanical properties of fracture healing in rabbits. *Clin Orthop* 1988;233:268-73.

Goldin, H et al. (1981). The effects of Diapulse on the healing of wounds: a double-blind randomized controlled trial in man. *British J of Plastic Surgery*, 34:267-70.

Goodman, R & Henderson, A. S. Sine waves enhance cellular transcription. *Bioelectromagnetics* 7:23-29; 1986.

Harrington, DB., Meyer. R & Klein, RM.: Effects of small amounts of electric current at The cellular level. *Ann. N.Y. Acad. Sci.* 238:300, 1974.

Harrington, DB.. and Becker. RO.: Electrical stimulation of RNA and protein synthesis in the frog erythrocyte. *Exp. Cell Res.* 76:95. 1973.

Hartshorne. On the causes and treatment of pseudoarthroses and especially that form of it sometimes called supernumary joint. *Am. J. Med. Sci.* 1:143; 1840.

Heffernan, M. (1995). The effect of a single cranial electrotherapy stimulation on multiple stress measures. *The Townsend Letter for Doctors and Patients.* 147:60-64.

Heffernan, M.: Comparative Effects of Microcurrent Stimulation on EEG Spectrum and Correlation Dimension, *Integrative and Behavioural Science*, July-September, 1996, Vol. 31, #3

Summary: 30 subjects were selected for a study comparing the effects of microcurrent on smoothing of EEG measurements of the brain. Subjects were randomly assigned to three groups – microcurrent (100uA) applied to earlobe, trapezius area of shoulder, and no stimulation. Electrodes were arranged so subjects could not tell which group they were in. Fast Fourier Transform (FFT) and correlation dimension from chaos analysis were used to measure results. The researcher found that microcurrent applied to the shoulders was markedly more effective in smoothing EEG patterns than earlobe or placebo. “This would represent a possible cost-effective alternative to neurofeedback in treating (anxiety and attention deficit disorders), by raising low regions in the FFT.

J.A. Spadaro, S.E. Chase, and D.A. Webster: Bacterial inhibition by electrical activation of percutaneous silver implants, *Journal of Biomedical Materials Research*, Vol. 20, 565-577 (1986)

Summary: Percutaneous silver wire implants were placed in rats, and the wounds inoculated with *Staphylococcus aureus* to test how much infection would spread. Microcurrent stimulation was passed through the wires, with + anodal current placed into implanted silver wire, and the – cathodal electrode placed on the rat’s belly as a ground. It was found that significant inhibition of infection occurred, with the most marked results at 20uA current level. “Metallic silver can be effectively and efficiently activated to elicit its anti-microbial activity by the application of microampere electrical current.”

reported by Lawrence Altman: Cell Channel Finding Earns Nobel Prize *New York Times Medical Science* section, October, 9, 1991

Summary: Two German scientists, Dr. Erwin Neher and Dr. Bert Sakmann, will share the \$1 million dollar Nobel prize for their development of the patch-clamp technique that allows the detection of minute electrical currents in cell membranes. This discovery, which “revolutionized modern biology” may shed light on the causes of several diseases, like diabetes and cystic fibrosis. This method allowed the detection of 20 to 40 types of ion channels that allow positive or negatively charged ions into and out of the cells. “This study confirmed that electrical activity is not limited to nerve and muscle tissue, as previously thought, but is intrinsic to ‘all kinds of other cells’”.

Jacobs JD, Norton LA. Electrical stimulation of osteogenesis in periodontal defects. *Clin Orthop & Rel Res* 1977;124:41-52.

Jahn, T. L. A possible mechanism for the effect of electrical potential on apatite formation in bone. *Clin. Orthop & Rel. Res.*56:261-273; 1968.

Jorgensen TE. The effects of electric current on the healing time of crural fractures. *Acta Orthop Scand* 1972;43:421-37.

Kohavi D, Pollack S, Brighton C. Short-term effect of guided bone regeneration and electrical stimulation on bone growth in a surgically modelled resorbed dog mandibular ridge. *Biomater Artif Cells Immobilization Biotechnol* 1992;20:131-8.

Kopczyk RA, Norton LA, Kohn MW. Bioelectric regeneration of bone in periodontal defects. *J Dent Res* 1975; 54:914-19.

Korostoff. E.: Stress generated potentials in bone: Relationship to piezoelectricity of collagen. *J. Biomech.* 10:41. 1977.

Kulig, K., Jarski, R et al. (1991). The effect of microcurrent stimulation on CPK and delayed onset muscle soreness. *Physical Therapy*, 71:6(supplement).

Lanyon. L. E.. and Hartman. W.: Strain related electrical potentials recorded in vitro and in vivo. *Calcif. Tissue Res.* 22:315.1977.

Lavine LS, Lustrin I, Shamos MH & Moss M. The influence of electric current on bone regeneration in vivo. *Acta Orthop Scan* 1971:42:305-314.

Lavine. L. S.. Lustfin, I & Shamos. M. H.: Treat C[~] cal Onnopaeocs and related Rescaeca men: of congenital pseudarthrosis of the tibia with direct current. *Clin. Orthop.* 124:69. 1977.

Liss, S. (1996). Neurochemical profiles following electrocranial stimulation. Presented at the Hans Selye Eighth International Conference on Stress. Montreux, Switzerland.

Lundeborg, TC, Eriksson, S.V & Malm, M. (1992) Electrical nerve stimulation improves healing of diabetic ulcers. *Annals of Plastic Surgery*, 29(4):328 31.

Manley Tehan, L, Microcurrent Therapy: Universal Treatment Techniques and Applications. Corona, CA: Manley & Associates; 1994

Martin, R. B & Gutman, W. The effect of electric fields on osteoporosis of disuse. *Calc. Tiss. Res.* 25:23-27; 1978.

Masureik C, Erikson C. Preliminary clinical evaluation of the effect of small electrical currents on the healing of jaw fractures. *Clin Orthop & Rel Res* 1977;124:84-91.

McClanahan, B. J.; Phillips, R. D. The influence of electric field exposure on bone growth and fracture repair in rats. *Bioelectromagnetics* 4:11-19; 1983.

Meyer, F.P., A. Nebrenski. 1983. Micro Stimulòation and placebo Effect Calif Health Review 2:1

Mercola, JM & Kirsch, D. The Basis for Microcurrent Electrical Therapy in Conventional Medical Practice, *J of Advancement in Medicine*, 1995;8(2): 83-97

Mitchell, P. 1976. Vectoral Chemistry and the molecular Mechanism of Chemiosmotic Coupling: Power Transmission by Proticity. *Biochem Soc. Trans* 4;400

Morgareidge, KR Chipman, MR, Microcurrent Therapy, *Physical Therapy Today*, Spring 1990:50-53

Nessler and Mass: Direct-Current Electrical Stimulation of Tendon Healing in Vitro Clinical Orthopedics and Related Research, April 1987

Summary: 80 tendons from white rabbits were surgically transected and removed from the animals after being surgically repaired. They were divided into 4 groups of 20, and cultured with 10 of each group being electrically stimulated, and half not. A 1.4 volt direct current connected through a 150 kOhm resistor was used for stimulation, at a current of about 7 uA. It was found that currents any higher than this caused discoloration of the tendons. Healing was measured by proline uptake and bridging of the repair site by the epitenon. Results: "a continuous direct current causes increased tendon cell activity within seven days and the increased activity may persist as long as 42 days." The researchers suggested that externally applied microcurrents may be preferable in future studies.

Mulder, G.D. (1991). Treatment of open-skin wounds with electric stimulation. Archives of Physical Medicine and Rehabilitation, 72:375-7.

Nessler, J.p., D.P. Mass. 1985 Direct Current Electrical Stimulation of Tendon Healing in Vitro. Clin Orthop 217:303-312

Nordenström BEW Biologically Closed Electric Circuits: Clinical, Experimental and Theoretical Evidence for an Additional Circulatory System Nordic Medical Publications Sweden 1983

Norton LA, Hanley K & Twrkeitz J. Bioelectric perturbations of bone. Angle Orthod 1984;54:73-87.

Norton LA, Rodan GA & Bourret LA. Epiphyseal cartilage cAMP changes produced by electrical and mechanical perturbations. Clin Orthop & Rel Res 1977;124:59-68.

Norton LA. In vivo bone growth in a controlled electric field. Ann NY Acad Sci 1975;238:466-77.

Norton, L. A.; Rodan, G. A. & Bourret, L Epiphyseal cartilage cAMP changes produced by electrical and mechanical perturbations. Clin. Orthop & Rel. Res. 124:59-68; 1977.

Noto, K., P. Grant. 1985. Comparative Study of Electro-Acuscope Neural Stimulation and Conventional Physical Therapy Modalities. Physical Therapy Forum 4:11

O'Connor BT, Charlton H, Currey J, Kirby DR & Woods C. Effects of electric current on bone in vivo. Nature 1969;222:162-67.

Oweye, L. N. Spieholz, et al.: Low-intensity Pulsed Galvanic Current and the Healing of Tenotomized Rat Achilles Tendons: Preliminary Report Using Load-to-Breaking Measurements Archives Physical Med Rehab, Vol. 68, 415-418 July 1987

Summary: 60 rats were divided into three groups of 20. One was unstimulated, one group had their Achilles tendons stimulated with positive (anodal) current, and the third group's tendons were stimulated with negative (cathodal) currents. A current of 75 microamps, at 10 Hz was used. Results: "The group treated with anodal current withstood significantly greater loads ($p < 0.001$) than did either the group which healed normally (i.e. without stimulation) or the group treated with cathodal currents".

Pilla, A. & Margules. G Dynamic interfacial electrochemical phenomena at living cell membranes: Application to the toad urinary bladder membrane system. J. Electrochem. Soc. 124:1697, 1977.

Pinkard JS. The effects of Electric Current on Condylar Cartilage and Bone Growth. Master Thesis, Case Western Reserve University, 1984.

Reichmanis, Marino, and Becker: Electrical Correlates of Acupuncture Points IEEE Transactions on Biomedical Engineering, November, 1975

Abstract: Employing a wheatstone bridge, skin conductance was measured over those putative acupuncture points on the large intestine and pericardium meridians lying between the metacarpophalangeal joints and the elbow. Results were compared to those from anatomically similar locations devoid of acupuncture points. "At most acupuncture points on most subjects, there were greater electrical conductance maxims than at control sites".

Richez, Chamay and Bieler, U. of Geneva: Bone Changes Due to Pulses of Direct Electric Microcurrent, Virchows Arch. Abt. A Path Anat. 357, 11-18 (1972)

Summary: 26 rabbits had platinum electrodes surgically implanted into the medullary cavities of their humerus bones. Microcurrent stimulation was applied at 50 and 250 uA, allowing pause periods of one second between one second treatment bursts. The scientists found that osteogenesis (bone growth) happened more around the cathode (negative polarity), and that slight tissue necrosis occurred around the anode. The tissues stimulated acted as capacitors, discharging 75% of the current absorbed during the rest periods. They concluded that pulsed current is superior to direct current for bone healing acceleration.

Rodan, GA.; Bourret, L. A.; Norton, L. A. DNA synthesis in cartilage cells is stimulated by oscillating electric fields. *Science* 199:690-692; 1978.

Rowley, McKenna, and Wolcott: Proceedings: Use of Low Level Electrical Current for Enhancement of Tissue Healing. Biomedical Scientific Instruments #10, 1974

Summary: This article is an overview of theory and research into the titled field.

Sansen W & De Dijcker, F.: The four-point probe technique to measure bio-impedances. *Electromyogr. Clin. Neurophysiol.* 16:509. 1976.

Schmukler, R.; Pilla, A. A. A transient impedance approach to nonfaradaic electrochemical kinetics at living cell membranes. *J. Electrochem. Soc.* 129:526-528; 1982.

Scott, J., R. Picker. A double Blind Study to Evaluate Muscle Strength in Athletes Treated with Electro-Myopulse. *Intl. Soc Electro-Acutherapy.* Feb. 27,1983

Shafer DM, Rogerson K, Norton L, Bennett J. The effect of electrical perturbation on osseointegration of titanium dental implants: a preliminary study. *Oral Maxillofac Surg* 1995;53:1063-8.

Shamos MH. & Layine LS.: Piezoelectricity as a fundamental property of biological tissues. *Nature* 213:267, 1967.

Shapiro E, Roeber FW, Klempner LS. Orthodontic movement using pulsating force-induced piezoelectricity. *Am J Orthod* 1975;76:251-254.

Sinitsyn, Razvozva (Russian): Effects of Electrical Microcurrents on Regeneration Processes in Skin Wounds *Ortop Travmatol Protez,* Feb. 1986

Summary: 68 patients with post burn and post traumatic wounds underwent treatment constant and modulated microcurrent of negative polarity of 1-10 uA/cm² over a period of 2-20 days. Although both groups showed accelerated regeneration, the modulated electric current group showed more prolonged and marked effect. Better survival of skin grafts was demonstrated compared with untreated patients.

Sinitsyn, Razvozova, (Russian): Stimulation of the Regeneration of Skin Wounds by Microcurrents Vopr Juroortol Fizioter Lech Fiz Kult, Nov.-Dec. 1985

Stanish and Gunlaughson: Electrical Energy and Soft-Tissue Injury Healing *Sportcare and Fitness,* Sept/Oct 1988

Summary: This article is a summary of research into tendon healing acceleration, including human injuries of the anterior cruciate ligament and the Achilles tendons: "While the results are subjective, the individuals in both groups appear to have returned to usual activities more quickly, and have greater mobility, than people treated more conventionally".

Stan. S, Muller, J; Sansen. W. & Dewaele. P.: Effect of direct current on the healing of fractures. In Burney. F., Herbst, E. & Hinsenkamp. M. (eds): *Electric Stimulation of Bone Growth and Repair.* Berlin. Heidelberg. New York, Springer-Verlag. 1978,pp. 47-53.

Stanish, W. Electrical Stimulation of Torn Ligaments Cuts Rehab Time by two-thirds. *Medical World News* Feb. 27, 1984, p. 67

Starlanyl, D *Fibromyalgia & Chronic Myofascial Pain Syndrome: A Survival Manual.* Oakland, CA: New Harbinger Publications, Inc.; 1996

Stefan A, Sanses W, Milier JC. Experimental study on electrical impedance on bone and the effect of direct on the healing of fractures. *Clin Orthop a Rel Res* 1976;120:264-67.

Steiner M & Ramp WK. Electrical stimulation of bone and its applications for endosseous dental implantation. *J Oral Implantol* 1990;16:20-7.

Stromberg, BV (1988). Effects of electrical currents on wound contraction. *Annals of Plastic Surgery*, 21:121-23.

Taubes, G. An Electrifyng Possibility. *Discover*. Apr. 1986, pp. 23-37

Travell, JG & Simons, DG Myofascial Pain and Dysfunction: The Trigger Point Manual, Vol. 1: The Upper Body. Baltimore, MD: Williams & Wilkins; 1983

Tomoya Ohno (Japanese): Experimental Studies of Influences on Healing Process of Mandibular Defect Stimulated by Microcurrent *Shikwa Gakuho*, #82 1982

Summary: 50 uA microcurrents were applied to one side of the jaws of a group of dogs with lesions in their jaws. The other side was untreated. The dogs were examined at periods of 3, 7, 14, 21, 28, 42 and 56 days. Results: "It seems likely that direct microcurrent promotes normal bone formation within the defective area and accelerates the osseous healing process. Prolonged application of electrical stimulus promotes a remarkable bone remodeling mechanism."

Vanable, Joseph: The Role of Endogenous Electrical Fields in Limb Regeneration Limb Development and Regeneration, Part A. pages 587-596 Alan Liss Publishing, N.Y. 1983

Watson, J. The electrical stimulation of bone healing. *Proc. IEEE* 67:1339-1351; 1979.

West, B.J. (1990). Fractal Physiology and Chaos in Medicine. World Scientific, New Jersey.

Weiss, D.S et al. (1990). Electrical stimulation and wound healing. *Archives of Dermatology*, 126:222-225.

Witt. H. T., Schlodder, E & Graber. P Membrane-bound ATP synthesis generated by an external electrical field. *FEBS Lett.* 69:272, 1976.

Wu. K. T., Go, N., Dennis, C., Enquist, I. & Sawyer P. N. Effects of Electric Currents and interfacial potentials on wound healing. *J Surg. Res.* 7:122. 1967.

Wolcott, Wheeler, Hardwicke, and Rowley: Accelerated Healing of Skin Ulcers by Electrotherapy *Southern Medical Journal*, July 1969.

Summary: These researchers applied microcurrent stimulation ranging from 200-800 uA to a wide variety of wounds, using negative polarity over the lesions in the initial phase, and then alternating positive and negative electrodes every three days. The treated group showed 200-350% faster healing rates than control, with stronger tensile strength of scar tissue and antibacterial effects in infected wounds in the treated group.

Zengo AN, Bassett CA, Proutzos G, Pawluk R, Pilla A. In vivo effects of direct current in the mandible. *J Dent Res* 1976; 58:383-90.